

Reinventing blood safety

Use of Radiolabeled Platelets for Assessment of In Vivo Viability of Platelet Products

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Desirable Platelet Radionuclide Tracer Characteristics

> Objective

- Internal or external quantitation of platelet kinetics.

> Radionuclide Characteristics

- Readily detectable.
- Physiologic element.
- Non toxic to cell/patient.
- No perturbation of study.
- Minimal reutilization/elution.
- Ease of administration/sampling.
- Selective tracer uptake.
- Homogeneous cellular distribution

Radionuclides - Principles & Practice

Principle

<u>Representative Dose</u>	<u>Stable Donor</u>	<u>Defined Distribution</u>	<u>Sample Precision</u>
Harvest Representative Aliquot	Variable Turnover	Estimated Volume	Constant Volume
No Selective Process Loss		Assumes Consistency	Consistent Sample Timing
Consistent Tracer Uptake	Variable Cell Quality	Affirm Steady State	
No Label Damage/Elution			Accurate Counting

Practice

43mL Whole Blood	Time Paired Studies	Nomogram Blood Volume Estimates	Weighed 2mL Samples
10- 20 ml Platelet Concentrate			
Tube Processing >80% recovery	Concurrent Studies	Dilute 3 Standards 1:5000	Correct for Injectate Plasma Elution
Uptake		5 -10% Immediate In-vitro Elution	Use 10 Day RBC Activity Correction
60- 80% ¹¹¹ In & 20- 40% ⁵¹ Cr		Platelet Counts	Count to 2% Error
1 X Soft + 2 X Hard Spins			

^{111}In & ^{51}Cr Labeling Characteristics

	<u>^{111}In-Oxine</u>	<u>^{51}Cr – Sodium Chromate</u>
Desirable Emission	90-94% @ 172 & 247 kev	9% @ 320 kev
Tissue Selective Uptake	Plt >> WBC >>> RBC	RBC >>> Plt >> WBC
Non Toxic (Target/Patient)	Oxine @ 3-6 ug/mL	Chromate < 10^4 molar
Detection Parameters	171 kev → ~72% eff 245 kev → ~53% eff	320 kev → ~3% eff
Elution (RBC & Plt)	8%/day & 11% @ day-1	1-2%/day & 6% @ day-1

^{111}In & ^{51}Cr Tracer Characteristics

	<u>^{111}In - Oxine</u>	<u>^{51}Cr – Sodium Chromate</u>
Administration Ease	↑ Transferrin Avidity (Wash)	Activity ≥ 20 uCi/ug Cr (Plt Count)
Clearance	Plasma T $\frac{1}{2}$ ~10 Hours	R/E → Excretion @ 3%/day
Reutilization	Nil post oxine ↓	Nil post chromate → Chromic
Cell Uptake	Cells Equivalent (80% cytosol)	Energy Dependent (ATP associated)
Half Life	2.8 days → rapid counting	28 days → delayed counting
Counting Technology	Correct for count times	3" crystal NaI detector= ~ 2x Eff.

Detection Implications of ^{111}In & ^{51}Cr

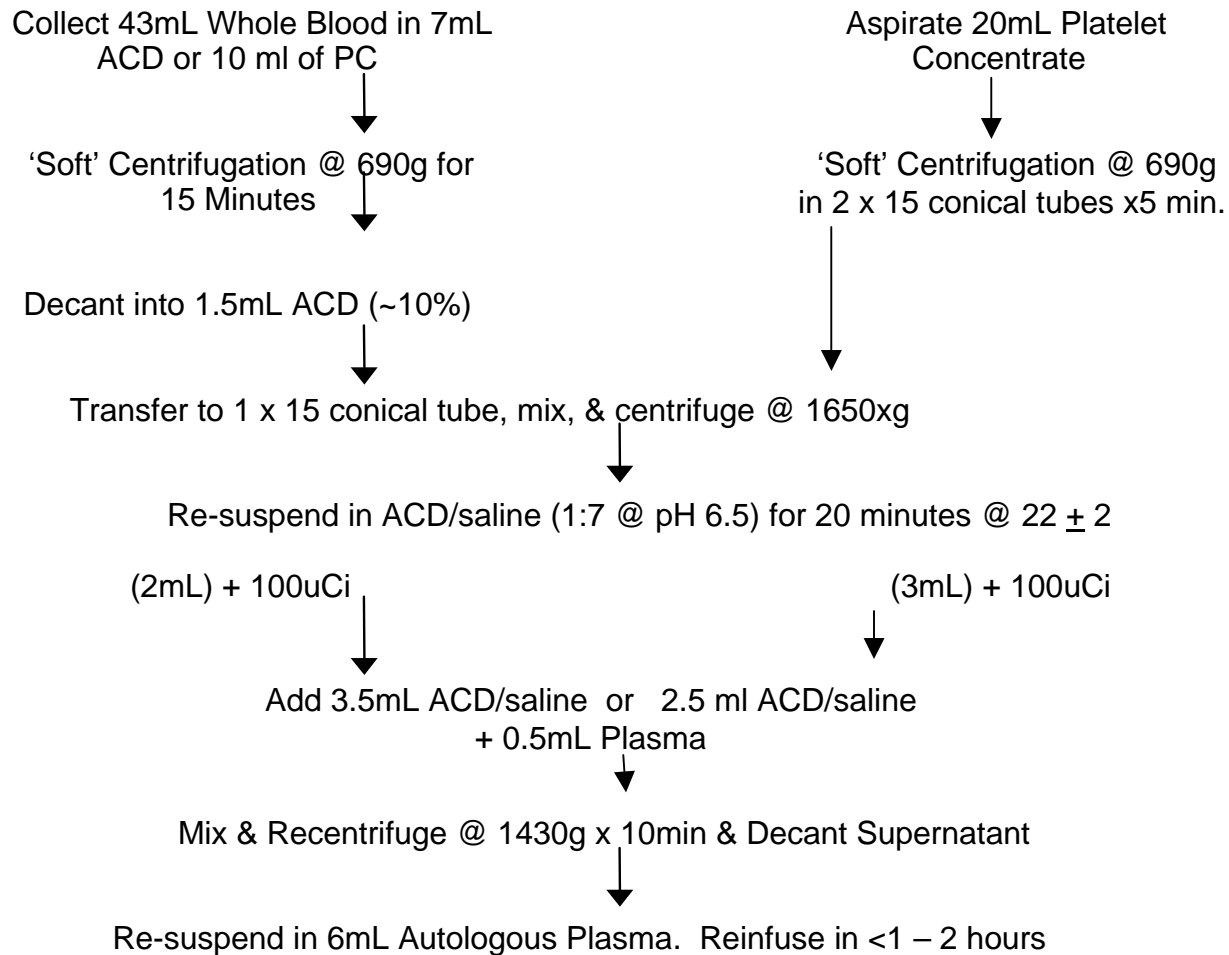
Physical Characteristics

- > Low ^{51}Cr photon yield mandates high efficiency (NaI) counters.
- > Photon scatter requires ^{111}In sum peak counting.
- > ^{51}Cr may be counted directly with scatter correction.
- > 28 day ^{51}Cr $T_{1/2}$ allows late counting post ^{111}In decay.
- > Low dose (low count) infusions need long count times.
- > 2.8 day ^{111}In $T_{1/2}$ requires elapsed time count correction.
- > Rapid post sample processing and counting is desirable.
- > Standard counts should be diluted to \approx sample counts.

Development of a Double Label $^{111}\text{In}/^{51}\text{Cr}$ Assay Method

- > Purpose
 - Develop comparable ^{111}In & ^{51}Cr platelet techniques for consistent results.
- > Study Plan
 - Evaluate relationship between in vivo & in vitro elution.
 - Evaluate ^{111}In labeling effects on platelet function.
 - Assess ^{111}In and ^{51}Cr RBC activity and evolution.
 - Develop corrections to support generation of equivalent outcomes.
- > Studies
 - Studies were performed using a similar tube/electrolyte method.
- > Procedure Development
 - Develop a procedure for simultaneous ^{111}In and ^{51}Cr platelet labeling.
 - Generate a simultaneous ^{111}In and ^{51}Cr infusion, sampling, and counting procedure.
 - Validate the essential equivalence of the two methods.

Double Isotope Platelet Procedure Development – Labeling Process



Development of a Double Label $^{111}\text{In}/^{51}\text{Cr}$ Assay Method

- > Elution (A)
 - 63 In vivo/in vitro studies performed.
 - Early injectate, diluted injectate, and in-vitro/in-vivo elution analysis.
 - Injectate processing method developed → injectate correction.
- > BioDistribution & RBC Elution (B)
 - 15 simultaneous ^{111}In and ^{51}Cr imaging and kinetic studies performed.
 - 0, 5, 10 day stored CPD-PC concentrates studies.
 - In vivo whole body and organ uptake measured over 24 hours.
 - RBC activity quantitated over 10 days → RBC correction.
- > $^{111}\text{In}/^{51}\text{Cr}$ Double Label Validation (C)
 - 16 concurrent ^{111}In & ^{51}Cr 5 day PC storage studies.
 - Post infusion platelet, RBC/WBC, & plasma activity density separated.
 - Double manual apheresis cross over study design developed.
 - Validation study to define sample size requirements.

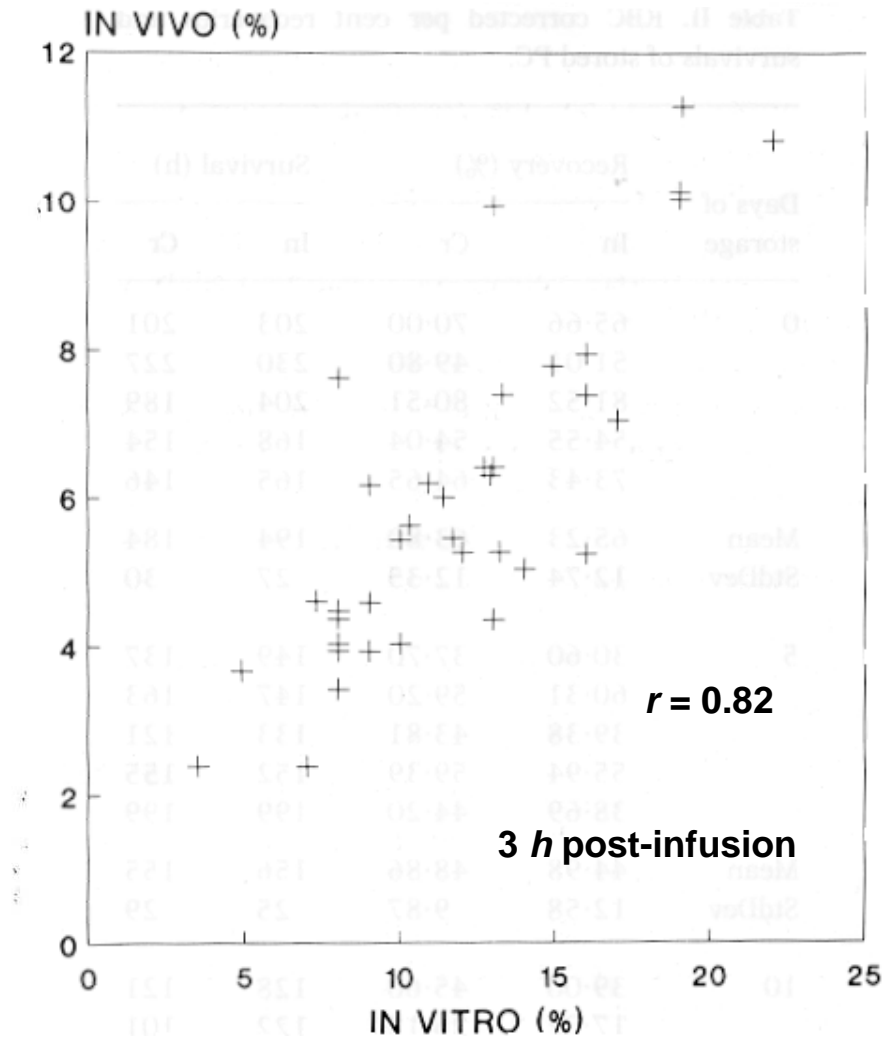
Double Label $^{111}\text{In}/^{51}\text{Cr}$ Development Studies

(Study A) In Vivo Plasma Activity (Mean \pm SD)

<u>Label Uptake</u>	72 \pm 8%	42 \pm 10%
<u>Labeling Loss</u>	35 \pm 9% (similar)	
<u>In-vivo - Post Tx Plasma Activity (%)</u>	<u>^{111}In</u>	<u>^{51}Cr</u>
5 Minutes	5 \pm 1	3 \pm 2
1 Hour	6 \pm 2	1 \pm .5
3 Hours	6 \pm 2	0.8 \pm 7
<u>In-vitro Plasma Activity</u>		
Neat Injectate (2 hours)	3 \pm 3%	6 \pm 3%
Diluted Injectate (2 hours)	* 11 + 4%	** 9 + 4%

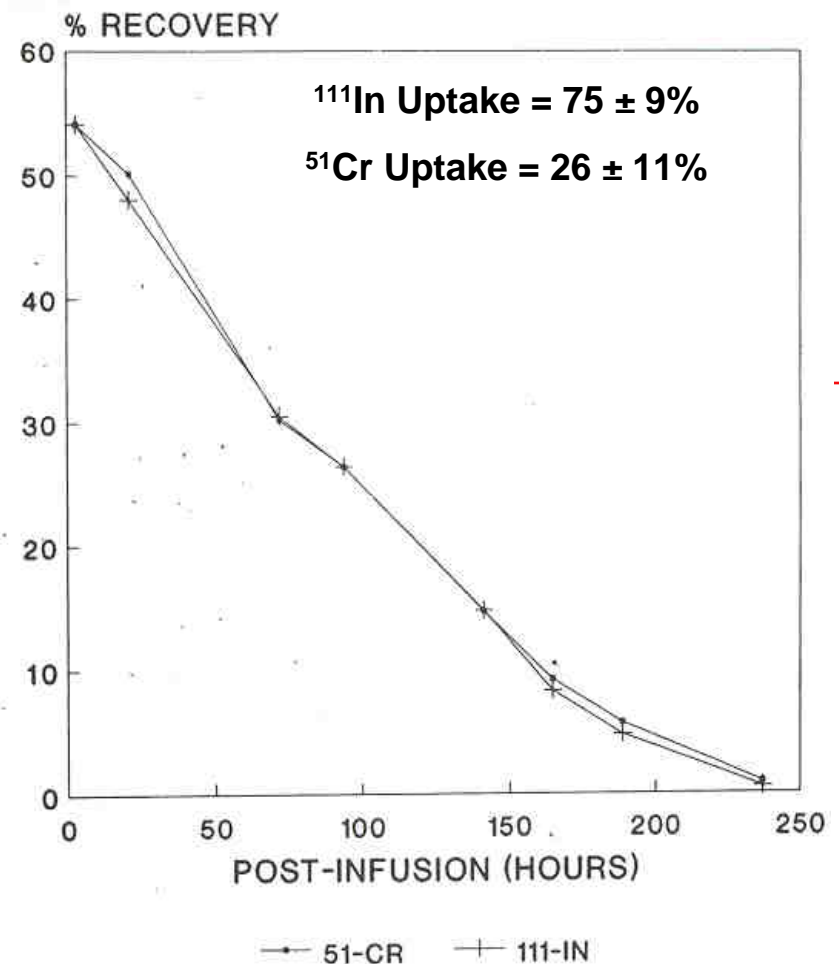
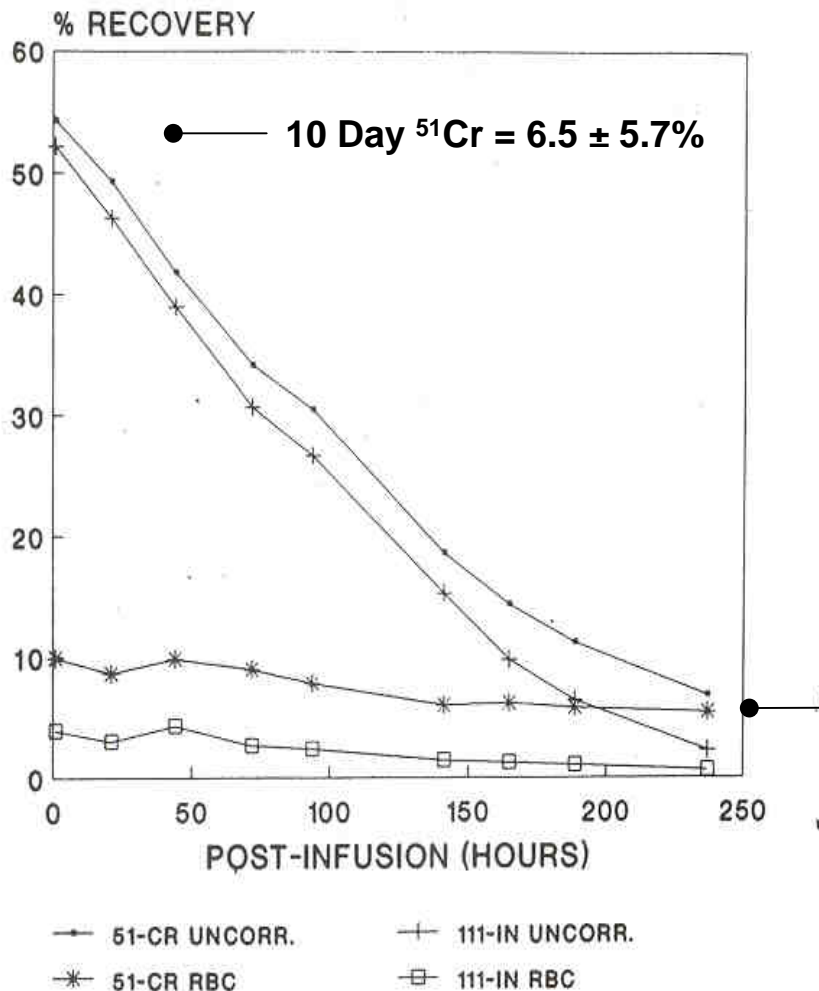
* In Vivo/In Vitro Correlation $r = 0.82$ * (@ 3 hours) $r = 0.77^{**}$ + (@ 5 minutes)

Comparison of In Vitro & In Vivo ^{111}In Activity



In vitro ^{111}In plasma activity of diluted injectate in fresh whole blood and *in vivo* plasma activity.

⁵¹Cr RBC Uncorrected & Corrected Recoveries



Double Isotope Platelet Procedure – Elution & Plasma Correction

> Injectate

- Retain aliquot of ^{111}In -only and ^{51}Cr -only injectate for standards.
- Mix ~ 15uCi of injectates and retain an aliquot for standards.
- At the time of infusion, add 10uL injectate to 10mL fresh EDTA blood.
 - Centrifuge after 2 hours @ 37 °C & calculate elution fraction.
- Prepare 3 x 1 in 5000 individual & mix injectate standards.

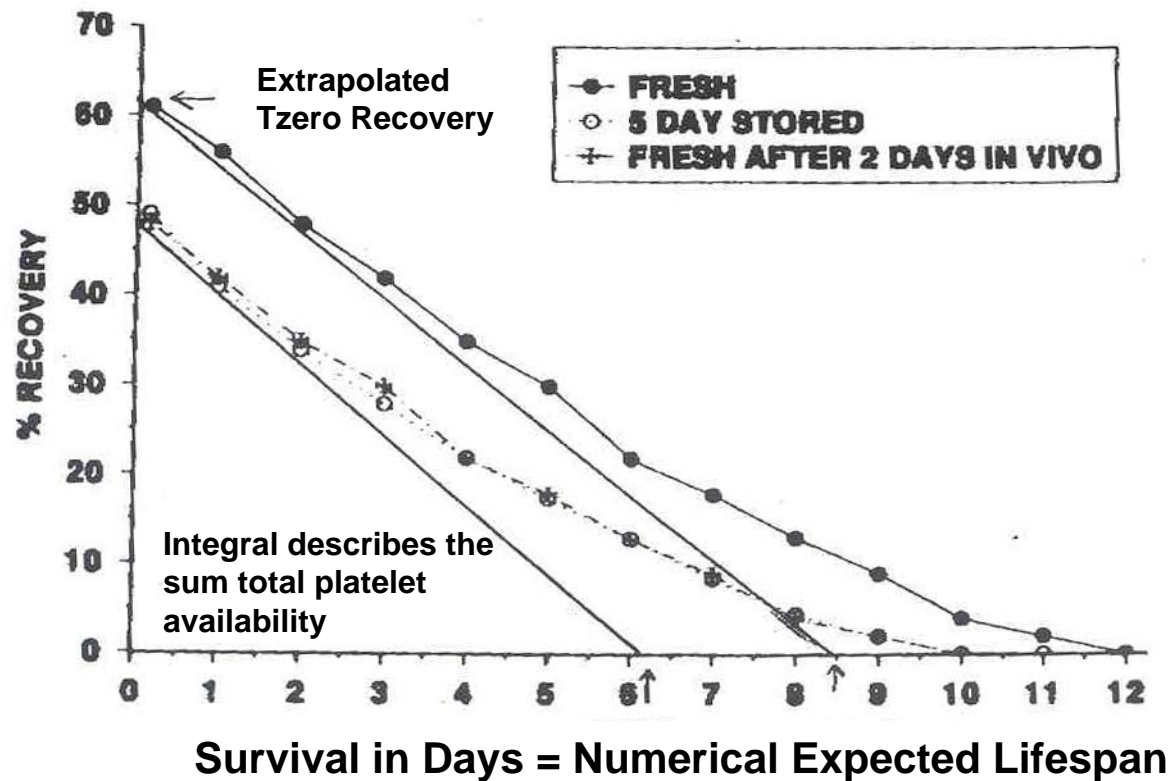
> Samples

- Collect 7mL EDTA samples @ 3 hours, 7 samples/10 days.
 - Prepare 2mL weighed aliquots for WB counting, & centrifuge an additional 2ml for split counting to correct for plasma activity.
- Count individual, and mix standards; samples; splits to a 2% accuracy.
 - Correct standards for elution and samples for RBC & plasma activity.

Corrected Post Transfusion ^{111}In & ^{51}Cr Platelet Kinetics

<u>Study B</u>	<u>Storage Duration</u>	<u>^{111}In</u>	<u>^{51}Cr</u>
% Recovery	0	65 ± 13	64 ± 12
	5	45 ± 13	49 ± 10
	10	24 ± 13	29 ± 12
Survival (Hours)	0	194 ± 27	184 ± 30
	5	156 ± 25	155 ± 29
	10	72 ± 53	63 ± 53
<hr/>			
<u>Study (C)</u>			
% Recovery	5	66.1 ± 10.6	65.6 ± 10.9
Survival (hours)	5	164.4 ± 25.5	164.4 ± 31.5
Integral (% hours)	5	6026 ± 1185	5958 ± 1240

Platelet In Vivo Kinetic Calculation Principles



Sample Size – Concurrent vs. Separate

% Recovery (absolute %)

Detection Goal	<u>10%</u>	<u>7.5%</u>	<u>5%</u>
Separate	16	22	32
Concurrent	5	6	8

Survival – (Hours)

Detection Goal	<u>30 Hours</u>	<u>25 Hours</u>	<u>20 Hours</u>
Separate	16	22	32
Concurrent	5	6	8

Table shows sample size required to detect a listed difference with 80% power and with $\alpha = 0.05$.

Storage Duration & ^{111}In Platelet Kinetics

> Purpose

- Evaluation of in vivo kinetics, storage duration, and in vitro parameters.

> Design

- 35 time separated paired ^{111}In platelet kinetic studies.
- Test (PAS) and control (CPDA-1) P.C. stored from 0.5 to 10 days @ 22 °C.
- ^{111}In studies performed with plasma correction.
- Post transfusion recoveries (PTR), survival (numerical expected lifespan), and integral (area under curve) were quantitated.
- Degree of exponential (random loss vs. senesce) estimated.
- Relationship between in vivo and in vitro parameters compared.
- In vitro measures included pH, HSR, ESC, ATP & lactate production.

Vox Sang 59:12:1990

Storage Duration & ¹¹¹In Platelet Kinetics

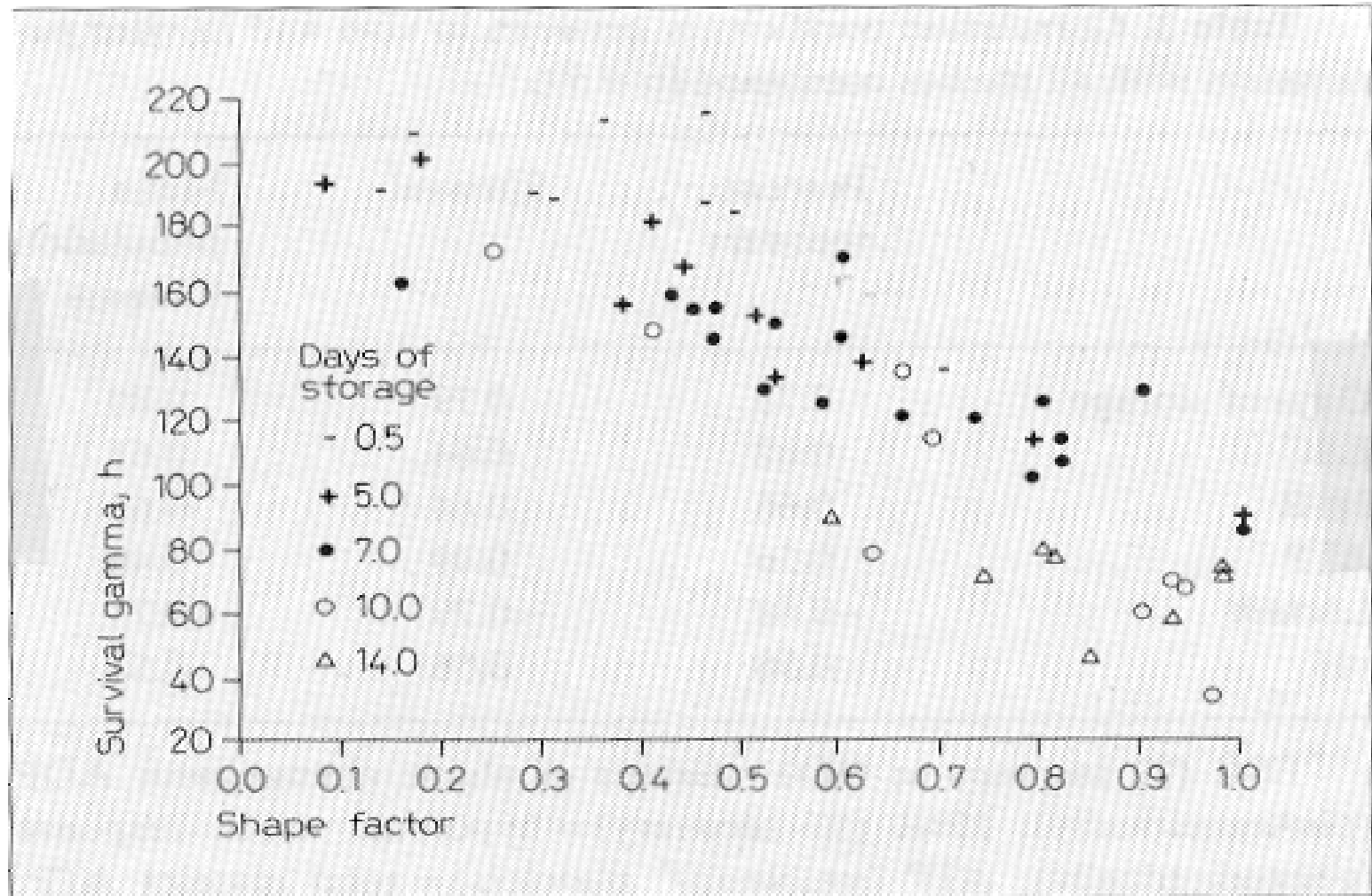
<u>Days Stored</u>	<u>% Recovery</u>	<u>Survival (Hours)</u>	<u>Shape Factor</u>
0.5	55 ± 10	189 ± 24	.38 ± .17
5	41 ± 11	146 ± 41	.55 ± .30
7	37 ± 11	107 ± 39	.83 ± .14
10	23 ± 9	74 ± 43	.87 ± .14
14	9 ± 8	51 ± 24	.88 ± .09

Implications

- Both PTR and survival decreased with storage duration.
- Donor variability mandated double label studies.
- Lactate, morphology, and pH independently correlated with ¹¹¹In kinetics.

Vox Sang 59:12:1990

Storage Duration & Non Linear Loss



Vox Sang 59:12:1990

Double Label ^{111}In Fresh & ^{51}Cr – Stored Platelet Study Design

- > Paired in vivo ^{51}Cr studies were performed ~ 28 days apart.
- > 18 whole blood donations were randomly processed into BC-PC or PRP-PC.
- > Following 5 days of 22 °C storage, ^{51}Cr in vivo studies performed.
- > Concurrent fresh ^{111}In and stored ^{51}Cr studies performed.
- > Test vs. control outcomes were compared.
- > Stored Cr values were expressed as a % of fresh In values to give Relative Recoveries and Survivals.
- > Platelet in-vitro studies included cell counts, pH, O₂ & glucose consumption, lactate production, ATP, morphology, HSR, ESC.
- > Platelet GP1b and LDH release rates were also measured.

Transfusion 32:113:1992

In Vivo Variables of fresh ^{111}In -labeled & 5-day stored ^{51}Cr -labeled PRP-PC* & BC-PC†

Platelet Recovery (%) ‡

	^{111}In <u>Fresh</u>	^{51}Cr <u>5-day-stored</u>	$^{51}\text{Cr}/^{111}\text{In}$ <u>relative (%)</u>
PRP-PC	60 ± 7	49 ± 10	81 ± 9
BC-PC	64 ± 6	53 ± 8	85 ± 10

Platelet Survival ‡

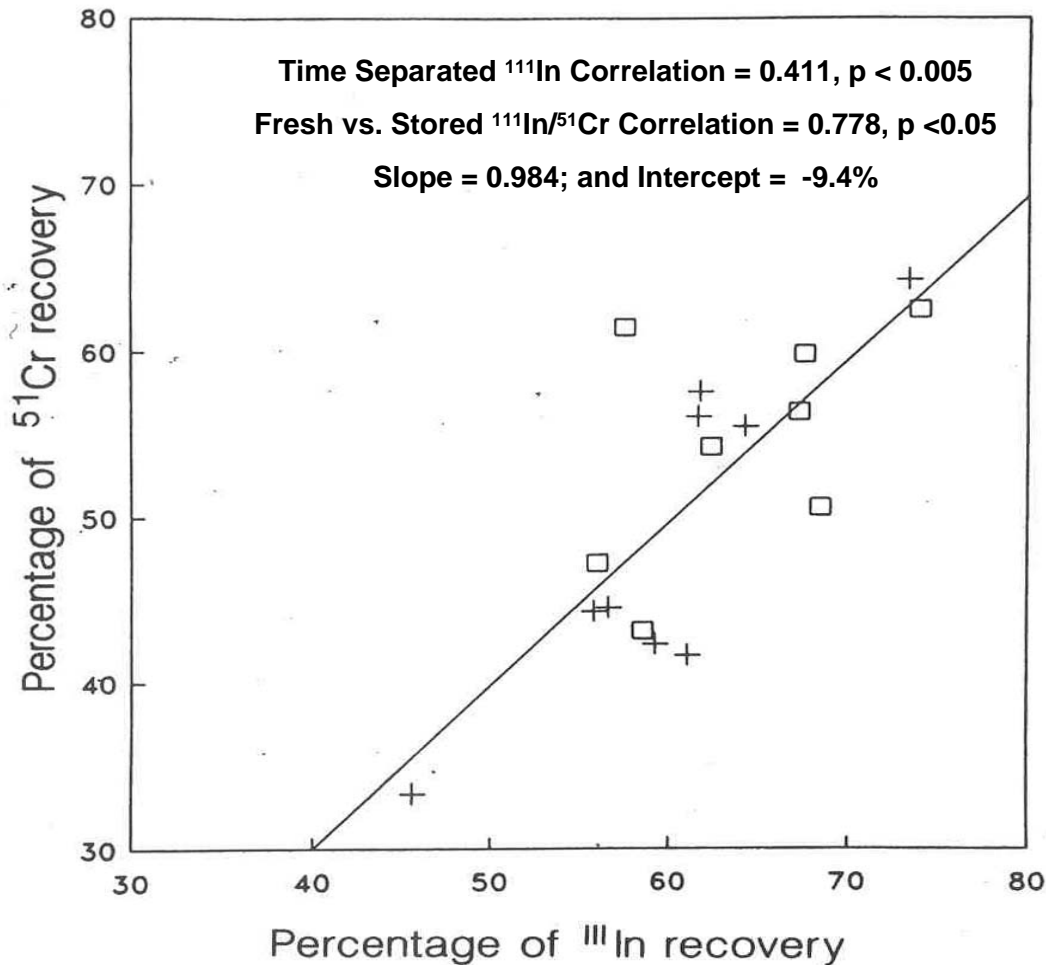
	^{111}In <u>Fresh</u> <u>(hrs)</u>	^{51}Cr <u>5-day-stored</u> <u>(hrs)</u>	$^{51}\text{Cr}/^{111}\text{In}$ <u>relative</u> <u>(%)</u>
PRP-PC	210 ± 22	162 ± 29	77 ± 10
BC-PC	209 ± 30	163 ± 20	79 ± 11

* Platelet-rich plasma-platelet concentrates.

† Buffy coat-PCs.

‡ Mean \pm SD.

Paired Concurrent Comparison of ^{111}In Fresh & ^{51}Cr Stored PC



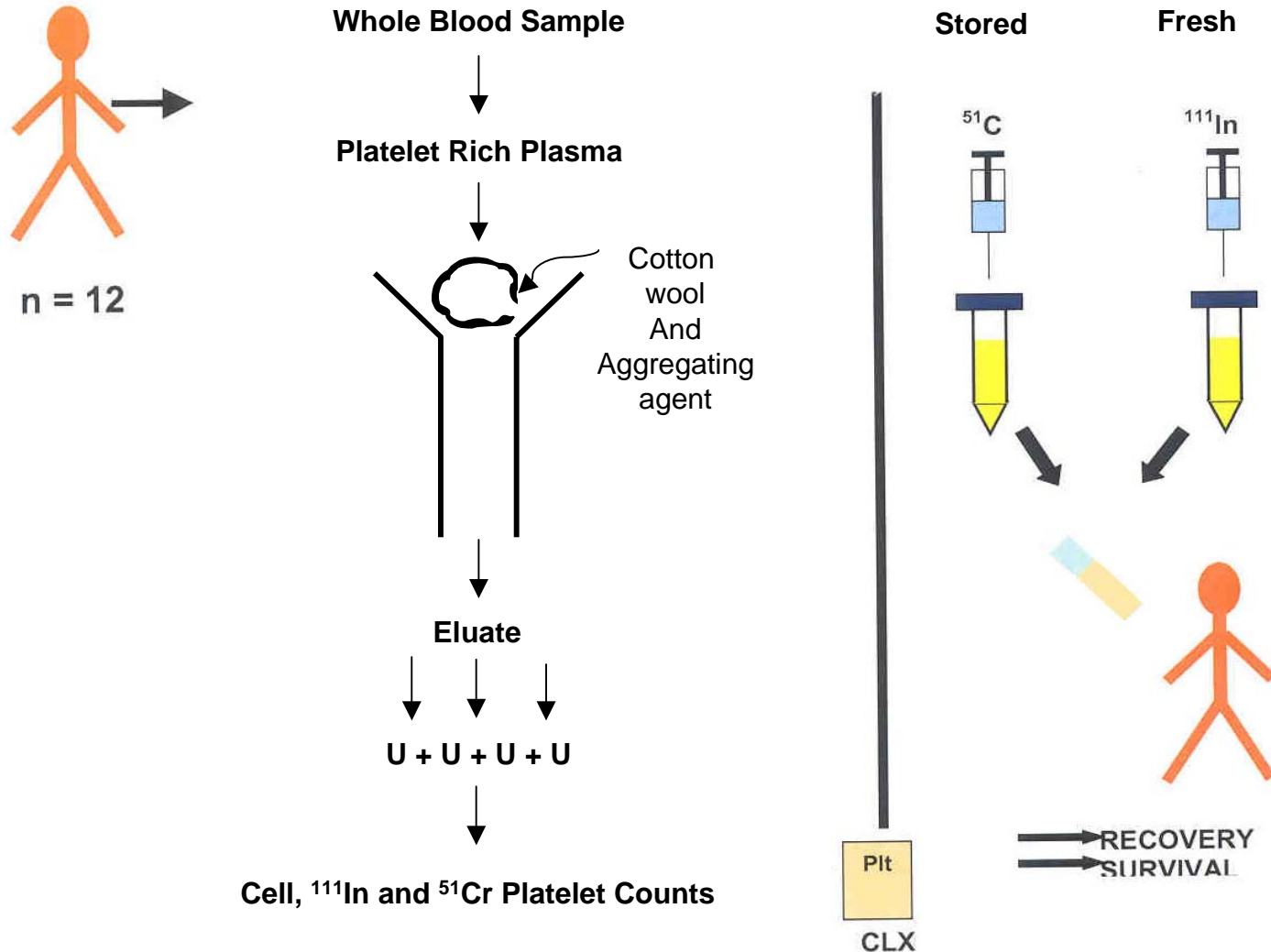
Simultaneous Paired ^{111}In & ^{51}Cr

Reduced Platelet Volume Study Design

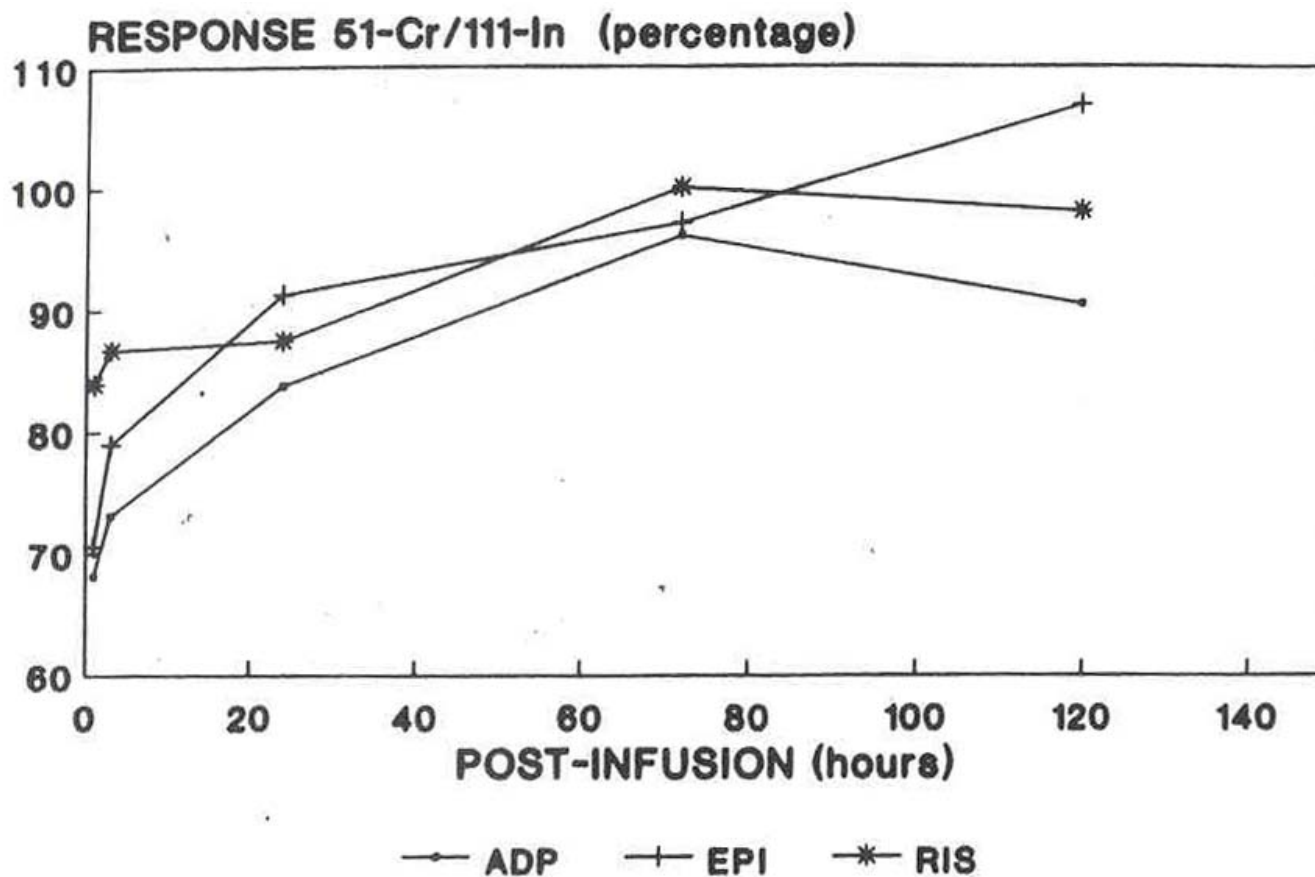
- > Apheresis into standard and reduced volume P.C.
- > 20 unit study with randomized order labeling with ^{111}In and ^{51}Cr .
- > Simultaneous infusion of ~ 15uC, ^{51}Cr and ^{111}In labeled platelets.
- > Elution and red cell correction applied.
- > Relative Recoveries (>35mL)
 - > % Recoveries = 99 (95 - 103)
 - > Integral (% .days) = 99 (96 – 101)

In Vivo Mean (95% CI) <u>Values</u>	<u>30 – 34mL P.C.</u>	<u>35 – 50mL P.C.</u>	
% Recovery	80 (69-92)	99 (95-103)	p = .005
Survival (days)	89 (83-94)	103 (98-107)	p = .0005
Integral (% days)	81 (69-93)	99 (96-101)	p = .012

In Vivo Post Transfusion Functional Recovery Study

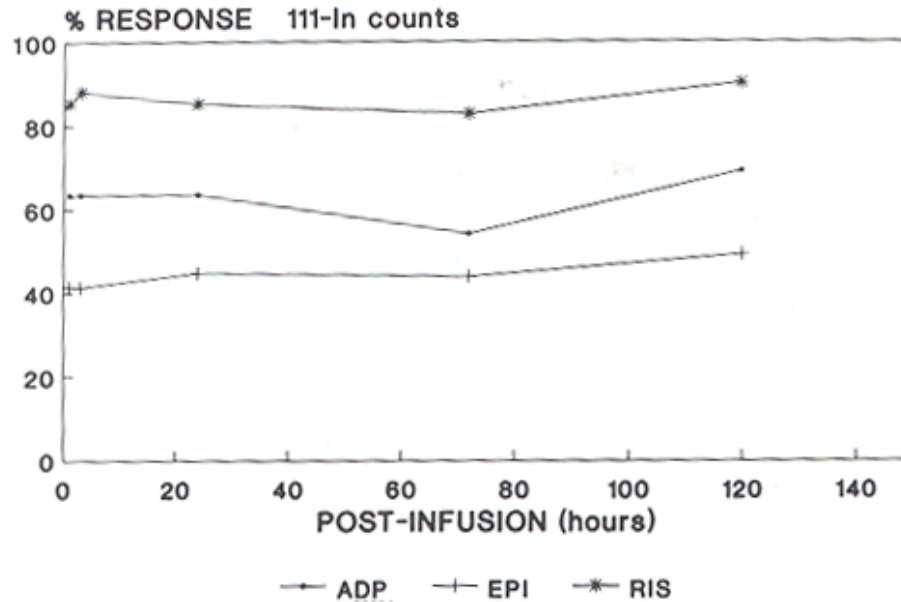


Ex-Vivo Aggregation of ^{51}Cr Stored Platelets



Percent ^{111}In labeled fresh platelet response

Post Transfusion Functionality of ^{111}In Platelets



^{111}In labeled fresh platelets as function of post-infusion time.

$N = 12$

No significant difference over 5 Days in vivo

^{111}In Aggregation = Numerical Aggregation

BJH 80:539:1992

Double Label ^{111}In & ^{51}Cr In Vivo Studies

- > Described a Double Label Method Identifying
 - Labeling issues relative to selective tracer uptake.
 - Technical issues relative to differential radionuclide counting.
 - Procedural issues relative to result acquisition/interpretation.
- > Reviewed Some of the Physiological Observations Associated with PC Storage
 - Storage associated loss of in vivo efficacy.
 - Sites of storage damaged platelet uptake.
 - Chronological variation in platelet turnover.
- > Proposed a Study Model to Allow Accurate Kinetic Analysis
 - Provided insight into P.C. functional recovery.
 - Suggested a driver to platelet senescence.